Standard Operating Procedure for:

Escherichia coli and Total Coliform using the **IDEXX** Quanti-Tray/2000 System with Colilert reagent

Ozarks Environmental and Water Resources Institute Missouri State University **Temple Hall 343** 901 South National Avenue Springfield, MO 65897

Prepared by: Brianns Cdwards OEWRI Research Specialist

Date: 09/22/2023

Approved by: <u>Robert Pavlowsky</u> OEWRI Director

Date: 09/22/2023

September 22, 2023





OEWRI-SOP-007

Table of Contents

Identification of the test method	. 3
Applicable matrix or matrices	. 3
Detection limit	. 3
Scope of the test method	. 3
Standard for <i>E. coli</i>	. 3
Summary of test method	. 4
Definitions	. 4
Interferences	. 5
Health and safety	. 5
Personnel qualifications	. 5
Reagents and standards	. 5
Sample collection, preservation, and storage procedure	. 6
Quality control	. 6
Calibration and standardization	. 6
Laboratory procedure	. 7
Data acquisition, calculations, and reporting	. 8
Computer hardware and software	. 8
Method performance	. 8
Pollution prevention	. 8
Data assessment and acceptable criteria for quality control measures	. 8
Corrective actions for out-of-control or unacceptable data	. 9
Waste management	. 9
References	. 9
Tables, diagrams, flowcharts and validation data	10
Appendix A	11
Total Coliform and <i>E. coli</i> Bench Sheet1	15

Identification of the test method

Escherichia coli (*E. coli*) using the IDEXX Quanti-Tray/2000 System with Colilert reagent (Standard Methods, 9223 B.)

Applicable matrix or matrices

This method is suitable for use with surface water samples.

Detection limit

The detection limit for this analysis is 1 Most Probable Number (MPN) per 100mL of sample.

Scope of the test method

This standard operating procedure describes the test method for the collection and analysis of water samples for the enumeration of *Escherichia coli* and total coliform bacteria.

Standard for E. coli

The U.S. Environmental Protection Agency (EPA) and the Missouri Department of Natural Resources (MDNR) have established specific water quality standards for recreational water contact. These standards apply between April 1st and October 31st and for water in losing streams year-round (MDNR 2022). These standards are as follows:

- For waters designated for whole body contact (WBC) Class A recreation during the recreational season or in losing streams at any time, the geometric mean (GM) for *E. coli* should be less than 126 per 100 milliliters (mL) of water (USEPA 1986, MDNR 2022).
- 2. For WBC-B designated waters, the GM value should be less than 548 per 100 mL (MDNR 2022).
- 3. Water designated for secondary contact recreation should not exceed 1,134 *E. coli* per 100 mL of water (MDNR 2022).

In addition to these standards, the EPA has defined a statistical threshold value (STV) for E. coli at 410 colony-forming units (CFU) per 100 mL of water (EPA 2012). This STV is akin to the 90th percentile, meaning it should not be exceeded by more than 10% of the samples taken. To assess compliance with these standards, the EPA recommends conducting weekly sampling and evaluating both the GM and the STV over a 30-day period or relating sampling frequency to the intensity of use of the water body (EPA 1986). Typically, E. coli data is presented in terms of CFU per 100 mL of water or most probable number (MPN), both signifying comparable measurements (MDNR 2014).

Summary of test method

Surface water samples are collected in EPA-accepted Screw-Top Sterile Coliform Polypropylene Water Sample Bottles. An undiluted sample will be analyzed from the sample collected. There will be a preliminary sampling event prior to project startup to allow the laboratory to determine if certain sites require dilutions. If a sample requires dilutions, a 1:10, 1:1 dilution, or both will be analyzed from the sample collected. The Colilert® reagent is added directly to the 100 mL undiluted sample in the Coli-test bag and to 100 mL of the diluted sample. Both are mixed thoroughly to dissolve the reagent. The samples are transferred to Quanti-Trays@/2000 and sealed using the Quanti-Tray sealer. Samples are incubated at $35.0 \pm 0.5^{\circ}$ C for 24 hours. Results are reported as MPN/100mL.

Definitions

- 1. Control cultures: For each order of Quanti Trays with different lot numbers, analytical procedures are checked by testing with known positive and negative control cultures. For example, *E. coli* is a positive control for this analysis and *Staphylococcus aureus* is a negative control.
- 2. Field duplicate (FD): Two samples taken at the same time and place under identical circumstances and that are treated identically throughout field and laboratory procedures. Analysis of field duplicates indicates the precision associated with sample collection, preservation, and storage as well as laboratory procedures.
- 3. Field blank (FB): An aliquot of deionized water treated as a sample in all aspects, including exposure to a sample bottle holding time, preservatives, and all preanalysis treatments. The purpose is to determine if the field or sample transporting procedures and environments have contaminated the sample.
- 4. Laboratory reagent blank (LRB): An aliquot of sterilized water treated as a sample in all aspects, except that it is not taken to the sampling site. The purpose is to determine if the analytes or interferences are present in the laboratory environment, the reagents, or the apparatus.
- Laboratory duplicate (LD): Two aliquots of the same environmental sample treated identically throughout a laboratory analytical procedure. Analysis of laboratory duplicates indicates precision associated with laboratory procedures but not with sample collection, preservation, or storage procedures.
- Relative Percent Difference (RPD): calculated as the difference between a sample and duplicate results, divided by the average of the sample and duplicate results, multiplied by 100%.
- 7. Method detection limit (MDL): The lowest level at which an analyte can be detected with 99 percent confidence that the analyte concentration is greater

than zero. The detection limit for this analysis is 1 Most Probable Number (MPN) per 100mL of sample.

Interferences

Water samples containing humic or other material may be colored. If there is background color, compare inoculated trays to a control tray containing only water.

Health and safety

- The analysis involves handling of freshwater samples that may contain live microorganisms and therefore pose some threat of infection. Laboratory personnel who are routinely exposed to such water samples are encouraged to protect themselves from water borne illnesses by wearing clean disposable gloves and washing their hands frequently.
- 2. The Colilert® reagent is not hazardous according to the manufacturer's material safety data sheet. The manufacturer does recommend wearing gloves and safety glasses while using this reagent and washing hands after use.

Personnel qualifications

Laboratory and field personnel shall have a working knowledge of this analytical procedure and will have received training from an MSU employee knowledgeable of the proper sample analysis procedures.

Equipment and supplies

- 1. Quanti-Tray Sealer®: catalog number WQTS2X-115. IDEXX Laboratories, Inc., Westbrook, ME
- Sealed Screw-Top Coliform Polypropylene Water Sample Bottle: 120 mL with 100mL fill line, sterilized and containing 10 mg of sodium thiosulfate. Thermo Fisher Scientific, 168 Third Avenue, Waltham, MA USA 02451.
- 3. Ultraviolet lamp, long wave for use in testing UV-florescent wells (6-watt, 365-nm UV light).

Reagents and standards

- 1. Colilert® reagent: for 100 mL samples, catalog number WP200. IDEXX Laboratories, Inc., Westbrook, ME.
- 2. Quanti-Tray®/2000: 100 trays containing 97 wells each, part number WQT-2K. IDEXX Laboratories, Inc., Westbrook, ME.
- 3. IDEXX Colilert® and Colilert-18 Quanti-Tray/2000 Comparator: catalog number WQT2KC. IDEXX Laboratories, Inc., Westbrook, ME.
- 4. Dilution water: sterile DI. Fisher Scientific, Biotech Grade Sterile Water, Cat. No. BP 2485-4, 4-liter PolyPac.

Sample collection, preservation, and storage procedure

- 1. Arrive at sampling site and record site number, date, and time.
- 2. Collect a 100 mL surface water sample by placing the sample bottle into the water. Lower bottle into the water, facing upstream, far enough so that the water flows freely into the bottle. Avoid capturing large particulate matter in the bottle.
- 3. The sample containers will be pre-labeled with a number representing the sampling sites. For each collection day, the date, stream flow and precipitation within the last 24 hours will be recorded in the field log book.
- 4. For each sample, the location number, bottle numbers used, and time of collection will also be recorded in the field sample log.
- 5. The samples will be kept in the possession of OEWRI personnel until transferred to the laboratory with appropriate chain of custody forms, where they become the laboratory's responsibility.
- 6. Samples will be transported to the laboratory in coolers containing ice. Transport should not take longer than three hours.
- 7. The samples will be stored at 4°C until analysis. The maximum hold time is 6 hours; however, they should be processed as soon as possible.

Quality control

- 1. Accuracy: Initial analyst demonstration of capability and for each new lot of Quanti-Tray/2000, analyze the following:
 - a. 1 replicate of a positive control organism
 - b. 1 replicate of a negative control organism
 - c. 1 replicate of sterilized water
- 2. Precision: the analyst should analyze:
 - a. Field duplicates (FD): two samples collected at the same time in the field to describe site variability.
 - b. Laboratory duplicates (LD): two counts are performed on positive wells to ensure counts are accurate.
 - c. Field blank (FB): Used to determine if the field or sample transporting procedures and environments have contaminated the sample.
 - d. Laboratory reagent blank (LRB): analyze one LRB per sample batch.

Calibration and standardization

There are no calibration or standardization procedures for this method.

Laboratory procedure

- 1. If the water sample bottle contains more than 100mL of water, decant the excess water to the 100mL fill line. The bottle containing the water sample should be shaken well just prior to preparation for analysis. For samples known to routinely go over range, prepare a 1:1 or 1:10 dilution.
 - a. If the sample was collected during <u>baseflow</u> and is expected to go over range, prepare a 1:1 dilution.
 - b. If the sample was collected during <u>stormflow</u>, assume sample will go over range and prepare a 1:10 dilution immediately.
- 2. Open a Colilert ampule and pour contents into sample bottle. Mix thoroughly and ensure the reagent is completely dissolved. Repeat for the remaining samples.
- 3. Follow manufacturer's instructions for preparation of Quanti-Tray/2000 and use of the Quanti-Tray Sealer (See Appendix A for the manufacturer's instructions).
- 4. Allow bubbles to settle or dissipate. Failure to do this may result in the wells filling or sealing improperly.
- 5. Record the sample's site code on the back of the tray for identification purposes.
- 6. Record the lot number of the reagents and the trays used on the bench sheet in the comments section.
- 7. Incubate at $35.0 \pm 0.5^{\circ}$ C for 24 hours. Record the time and incubation temperature at the start and end of incubation.
- 8. Count the number of small and large positive wells that are yellow, equal to or greater than the comparator, and refer to the MPN table to find the most probable number for total coliforms. The Colilert Comparator can be used to distinguish a minimal positive from a negative test result.
 - a. If well is yellow, but lighter than Colilert Comparator, incubate 4 additional hours and reread results.
 - b. Any intensity of yellow color equal to or greater than the comparator is considered a positive test result.
 - c. If sample remains less yellow than the comparator, it is negative for total coliforms and *E coli*.
- 9. Check <u>vellow wells</u>, both large and small, for presence of fluorescence by placing the wells under a blacklight. A well that fluoresces but is not yellow should **not** be marked as an *E*. coli positive well. Refer to the MPN table to determine the *E*. coli concentration.
 - a. If the yellow well fluoresces, but less than Colilert Comparator, the well is considered negative.
 - b. If the yellow well fluoresces equal to or greater than Comparator, the well is positive.
- 10. Record results on the bench sheet.

11. The completed bench sheet should be reviewed by the analyst, the laboratory manager, and the OEWRI QA coordinator.

Data acquisition, calculations, and reporting

For each sample analyzed, including quality control samples, record the number of small and large positive wells and the MPN in the appropriate places on the bench sheet (see below). Calculate precision for duplicate analyses using equation 1.

Equation 1. Relative Percent Difference (RPD) =
$$\begin{pmatrix} (A - B) \\ (A + B)/2 \end{pmatrix}$$
 100
Where: A = original sample MPN
B = duplicate sample MPN

For samples for which dilution was required, the concentration in the original water sample is calculated using Equation 2.

Equation 2: MPN_{sample} = MPN_{analysis} X (10.0 mL/V_{aliquot}) Where: MPN_{sample} = the concentration in the original water sample MPN_{analysis} = the concentration of the solution as determined on bench sheet V_{aliquot} = the volume of the aliquot diluted to 10 mL

Computer hardware and software

IDEXX MPN Generator 3.2: For use in confirming MPN of sample results, as well as calculation of 95% confidence intervals.

Method performance

There are no published method performance data for this method.

Pollution prevention

All waste from these procedures shall be collected and disposed of according to existing waste policies within the MSU Biology Department. Volumes of reagents made should mirror the number of samples being analyzed. These adjustments should be made to reduce waste.

Data assessment and acceptable criteria for quality control measures

- 1. The analyst should review all data for correctness (e.g., use of MPN table).
- Precision values are calculated for pairs of duplicate analyses. Record the precision values as a percent on the bench sheet. The desired precision is ± 20%.

- 3. The desired detection limit is 1 MPN/100mL.
- The completed bench sheet is reviewed by the analyst's supervisor or the OEWRI QA coordinator. Using IDEXX MPN Calculator 3.2 MPN, results are compared, and 95% confidence intervals are generated.

Corrective actions for out-of-control or unacceptable data

- 1. The results for precision and blank data are compared to the acceptable values for this analysis; ± 20% and 1 MPN/100mL, respectively.
- If a precision value exceeds 20% then the analyst should write in the comments section of the bench sheet: "These data are associated with an out-of-control duplicate analysis. The UCL = 20%." Note: "UCL" is the Upper Control Limit (i.e., 20%).
- 3. If a blank value exceeds 1 MPN/100mL then the analyst should write in the comments section of the bench sheet: "These data are associated with a blank value that exceeds the detection limit of 1 MPN/100mL."
- 4. The samples cannot be reanalyzed because the sample volume will be depleted after the initial analysis.
- 5. If data are unacceptable for any reason, the analyst should review their analytical technique prior to conducting this analysis again.

Waste management

The waste generated by this method is not hazardous. Waste can be discarded in the following manner: the water can be discarded in the laboratory sink and Quanti-Trays are autoclaved and then can be discarded with the paper trash.

References

- 1. EPA, Environmental Protection Agency. Water Quality Standards for Coastal and Great Lakes Recreation Waters. 2004.
- EPA, Environmental Protection Agency. Implementation Guidance for Ambient Water Quality Criteria for Bacteria - Office of Water. Washington, D.C., USA. 1986.
- 3. EPA, Environmental Protection Agency. Recreational Water Quality Criteria Office of Water 1.2.1. Washington, D.C., USA. 2012.
- 4. IDEXX Laboratories, Inc. Westbrook, ME 04092. Instruction manuals for use of: Colilert®, Quanti-Tray®/2000, and Quanti-Tray Sealer®.
- MDNR, Missouri Department of Natural Resources. Rules of Department of Natural Resources. Division 20 Clean Water Commission. Chapter 7 – Water Quality. 2022. 10 C.S.R. § 20-7.031.
- 6. MDNR, Missouri Department of Natural Resources. Water Protection Program Fact Sheet, PUB2401. 2014.

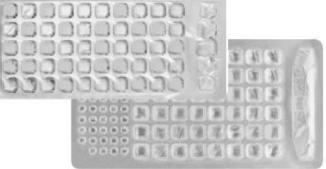
- 7. MDNR, Missouri Department of Natural Resources. *E.Coli* Monitoring at the Lake of the Ozarks, Environmental Services Program/Water Protection Program fact sheet. 2011.
- 8. Standard Methods for the Examination of Water and Wastewater. Method 9223 B., APHA, 21st Edition. 2005.

Tables, diagrams, flowcharts and validation data

- 1. See Appendix A for MPN tables and Quanti-Tray/2000 instructions.
- 2. See below for the bench sheet. The analyst should make a copy of this form for each batch of samples analyzed.

Appendix A





Quanti-Tray Certificate of Sterility

This certifies that the enclosed Quanti-Trays have been sterilized with ethylene oxide. For further information or documentation, contact IDEXX Laboratories, Inc.

> IDEXX Laboratories One IDEXX Drive, Westbrook, Maine 04092 USA

> > Phone 1-800-321-0207 Fax 207-856-0630

06-02320-07

Quanti-Tray®/2000

Introduction

IDEXX Quanti-Tray/2000 is designed to give quantitated bacterial counts of 100 ml samples using IDEXX Defined Substrate Technology* reagent products. Add the reagent/sample mixture to a Quanti-Tray/2000, seal it in a Quanti-Tray Sealer and incubate per the reagent directions. Then count the number of positive large and small wells and use the MPN table attached to determine the Most Probable Number (MPN).

Contents

This package contains 100 sterile Quanti-Tray/2000s.

User Instructions



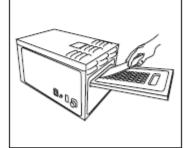
 Use one hand to hold a Quanti-Tray upright with the well side facing the palm.



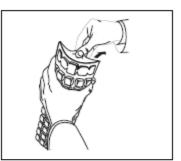
 Pour the reagent/sample mixture directly into the Quanti-Tray avoiding contact with the foil tab. Tap the small wells 2-3 times to release any air bubbles. Allow foam to settle.



 Squeeze the upper part of the Quanti-Tray so that the Quanti-Tray bends towards the palm.



 Place the sample-filled Quanti-Tray onto the Quanti-Tray/ 2000 rubber insert of the Quanti-Tray Sealer with the well side (plastic) of the Quanti-Tray facing down.



- Gently pull foil tab to separate the foil from the tray. Avoid touching the inside of the foil or tray.
- Seal according to Sealer instructions.
- Incubate according to reagent directions.
- Count large and small positive wells and refer to the Quanti-Tray/2000 MPN table to find the Most Probable Number (MPN).
- Dispose of media in accordance with Good Laboratory Practices.

For Technical Assistance, visit www.idexx.com/water, or in the U.S. and Canada, call 1-800-321-0207 or 1-207-856-0496.

IDEXX Laboratories, Inc. One IDEXX Drive, Westbrook, Maine 04092 USA

*Quanti-Tray and Defined Substrate Technology are either trademarks or registered trademarks of IDEXX Laboratories, Inc. in the United States and/or other countries. Covered by U.S. Patent Numbers 4.925,789 ; 5,429,933 ; 5,518,892. Other patents pending.



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X		7	22	5 6	103	11.4	12.6	13.8	15.0	16.3	17.6	18.9	202	21.6	230	259	27.5	29.1	30.7	32.4	34.1	35.9	37.7	39.7	41.7	43.7	45.9	10 10	100	224	58.1	609	63.8	67.0	73.8	77.6	81.6	86.0	959	101.7	108.1	115.3	123.6	133.3	145.0	159.7	179.3	in the United States
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		5	20	0 F	80	9.3	10.5	11.6	12.8	1	15.3	16.6	17.9	18.3	8 8	222	0.8	26.6	28.1	29.8	31.5	33.2	800	88	38.8	80.8	0 0 1		0.0		54.6	67.3	00.2	58	80.7	73.3	1.1	8.9	9.00	0.96	6 10	108.6	116.2	125.0		148.3	0.89	X Laboratories, Inc.
		4	9	2.5	12	8.3	9.4	10.6	11.8	13.0						223		25.3	26.9							39.3	4	0 F						613		712	74.9	78.9				105.4 1						
		69	30	3 2	55	72	8.4	9.5	10.7	6	13.1			16.9		34.1		24.1	25.6									33				53.8		88			727		822			102.2					1523	trademar
		2	5.0	0.5 F ¥	5	6.2	7.3	8.4	9.6	10.8	12.0	13.2	12	201	1.71	0 g			24.3						9	8.4		4 4 8 4	0 0	1				9.6			20.6	74.4				5.8	_				146.4	a registered trademark of IDE
		1	23				6.3	7.4	8.5						190			21.6	23.1									R 0 7						55.7			68.4	722				96.1					140.8	nark or a
		0		2 8					7.5				21			191			21.8			282						4 4						683			6 83	82									136.5	er a trader
eß	lls	ive										-					-			.,												•					-			~	~			-	-	-	-	"Quanti-Tray is either a trademark or
# Large	Wells	Positive	0		. 0	4	6	ø	7	00	ø	¥	ŧ	1	# :	λr έτ	*	4	*	*	я	74	8	8	两	8	स्य	218	48	9.8	ह	19 19	8	स १	\$ 18	8	8	84	4 4	4	4	4	*	*	đ	4	4	Cuanti-T

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		4		512									5 6					6,16		8.7	33.8						1162		130.8				16.4.4		183.3	194.7			241.1								6 >2419.6
		4	48.5	8	2.8	8	6.8	58.7	8,6	62.6	64.6	8	9 0 7 8	2 22	191	78.2	8.8	83.5	8.8	87	92.2						142		128.6		-				179.9				2360					-		960.6	3 2419.6
		\$	47.4	8	88	200	658	57.6	884	61.4	63.4	84	0.20	220	74.4	76.8			848	87.6	90.7	838	52.1				1123					144.0			176.6	187.3			2112					L.		913.9	1986.3
		\$	46.3	6.79	0 0 7 7	5.0	54.6	8	5.83	80.2	82.1	64.2	8.8	202	73.0	76.4	77.9	80.5	83.3	8	89.1	82.2	8.5	98.9	102.5	281	110.4	100	124.2	120.5	135.3	141.5	9 F	164.0	173.3	183.7	186.7	209.6	0.022		2012	100		483.3	616.7	870.4	1732.9
		4	45.3	46.8	4 0 4	517	53.5	552	57.1	59.0	609	629	67.1	693	71.6	74.0	76.5	79.1	81.8	84.6	87.5	906	93.8	97.2	100.7	0.40	108.5	5.24	12	127.3	132.9	139.0	0.041	161.0	170.0	180.2	191.8	206.3	2000	1047	8566	0.000	6788	487.4	593.8	829.7	1553.1
		\$	442	19	0.14	8	52.3	2	638	57.7	202	61.7	38	680	203	72.6	75.1	77.6	80.3	8	829	0.68	<u>82</u> .1	85.5	000	102/	106.6	15.00	1200	125.1	130.5	138.5		158.0	166.8	176.7	188.0	2010	216.4		0.107	2007	376.2	\$52.0	571.7		1413.6
		42	4	4.7		9 9 9	51.2	52.9	54.7	58:5	58.4		677.4 674.5		6.89	4.0	73.7	76.2	78.8	81.5	84.4	87.4	8.5	83.8	97.2	B	104.7	0.001	117.8	12.8	128.2	134.0		180	163.6	173.2	184.2	196.8	7.02	e e	0.102	345.4	384.9	437.1	550.4		1299.7
		ŧ	42.1	43.6	104	483	50.0	51.7	53.5	55.3	57.2	592	210	654	67.6	669	723	74.8	77.3	80.0	82.8	85.8	88.9	92.1	95.5	1.88	1029	8 F	115.7	120.6	125.9	131.6	0.701	152.1	160.5	109.8	180.4	1827	2040	1 2 2 2 2 2	1047	3063	353.8	4225	529.8	721.5	1203.3
ble	2	4	41.0	52	ġ ś	10	6.84	808	523	54.1	88	6/9		641	683	68.5	607	73.3	75.9	78.5	81.3	842	87.2	90.4	88	5/16	101.0	ŝ	113.7	118.5	123.6	129.2	200	149.2	157.3	108.5	176.8	188.7	2023	1.812	2846	0.007	385	408.3	6,603	689.3	1119.9
N Ta		8	6.0	4 (4		8	57.7	40.4	5	0.83	54.8	28	e ⊳ 8 8	8.29	6,49	67.2	8.5	49	74.4	20	79.8	82.6	8.8	88.7	82.0	ŝ	85	3 6	1118	116.3	121.4	128.8	8	146.4	154.3	183.1	173.2	184.7	1881	1 2 2 2	1 190	1.02	32.5	345	490.7	658.6	1046.2
MPI		8	38.9	40.4	5 1 A	450	46.6	48.3	50.0	51.8	53.6	56.5	104	615	63.6	65.8	68.1	70.5	73.0	75.5	78.2	81.1	84.0	87.1	90.3	1.55	97.3	1 2 2	185	14.2	119.1	124.5	2001	143.6	151.3	159.9	160.6	180.7	200	5.60	2510	281.0	3223	381.1	472.1	629.4	980.4
000	sitive	37	37.8	883	ŝ	9 8	45.5	5	48.8	50.6	52.4	8	8 8	602	62.3	8,89	68.7	8	71.5	74.1	76.7	79.5	82.4	88.4	886	220	88	e e e e e e e e e e e e e e e e e e e	1075	112.0	116.9	121	0/21	140.8	148.3	156.7	1881	176.9	204.2		2445	CM0	312.3	- <u>8</u> 8	454.1	01.5	8,028
v */2	IIs Po	8	8.8	83	R R	1 8 6	4.44	48.0	1.14	49.4	51.2	ŝ	n a 8 8	3	6.09	8	66.3	67.7	27	72.6	75.2	611	8.08	8.8	68	8	88	4 6 6 6 6	185	6.00	114.7	119.8	8	138.0	145.3	153.5	162.6	121	2 ş	n ar	1.017		30.6	385.5	436.6	574.8	866.4
Trav	Small Wells	8	36.7	37.2	202	418	432	44.8	46.5	48.2	50.0	51.8	100	57.6	59.6	61.8	64.0	66.3	68.6	717	73.7	76.4	792	822	852	000	91.9		1005	107.8	112.5	117.5	128	136.3	142.4	150.3	1592	187	101.1		2318	2581.0	2833	343.3	419.8	549.3	816.4
nti-	# Sma		34.7	8	9.6	82	4	43.7	63	47.0	48.8	88	672		583	60.4	62.6	649	67.2	60.7	72.2	74.9	77.6	80.5	83.6	8	5 8	1	101.5	106.7	110.3	115.2		132.6	139.5	147.3	156.9	1 <u>8</u>	17.0	2000	226.8	200	284.1	31.4	403.4	524.7	70.1
Quanti-Trav */2000 MPN Table		8	33.6	8	n o R b	8	9	42.6	4	42.9	41 B	83	7 5	3	0.72	58	612	83.5	8.8	68.2	70.7	73.3	78.1	78.9	819	8	8 9	n 4 5 8	88	103.7	108.2	113.0	7011	129.9	136.7	1442	152.6	162.1	1730	2011	1.102	2436	2753	319.9	387.7	5012	7.27.0
IDEXX		32	32.6	340	4 8 8 8	88	868	414	430	44.7	48.4	8	R 8 2	2.28	198	57.8	663	53	64.4	68.8	69.2	71.8	74.5	77.3	803	212	988	22	97.5	101.6	106.0	110.7	1213	127.3	133.9	1412	140.4	158.6	108.1	100	2140	2367	208.7	308.8	372.5	478.6	636.7
D		5	31.5	32.9	n y	37.2	38.7	403	6 ¥	43.5	62	69	- e 6	525	54.4	884	58.5	60.7	63.0	66.3	67.7	70.3	72.9	78.7	78.6	1.10	848	88	38	808	103.9	18.5	0.511	124.7	131.1	138.2	148.2	<u>8</u>	8	101 3	208.4	2002	258.4	208.1	357.8	456.9	643.8
		8	30.5	339	8 2	1	37.6	30.2	6	42.3	410	22	0 6 0 8	2 2	8	18	57.2	83	61.6	639	88.3	8.8	44	74.1	0.17	8	5 8 5 8	e a 8 8	8	97.6	101.8	106.3	7 11 7	122.2	128.4	135.3	143.0	151.7	161.5	1/3/0	0.001	2036	2204	287.8	343.6	436.0	613.1
		8	29.5	308	272	8	36.5	38.0	396	412	42.8	45	5 67 7 1 1 1	66	51.8	53.8	898	68.0	0 02	62.4	64.8	67.3	808	72.5	75.3	/93	81.4 exe	8 F	617	928	69.7	1042		119.6	125.7	132.4	130.9	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	15/ 2		8.101	0712	2027	277.8	330.0	416.0	579.4
		8	28.4	808	1.16	300	8.4	8.8	39.4	40.0	41.6	43.3	0 8 6 8	9.9	20.5	82.5	54.5	8.8	58.8	61.0	63.3	8.8	68.3	71.0	73.7	10.0	1.6	n 19		8.8	97.7	180	8	117.1	123.0	129.6	136.8	150	104.2		190 4	2410	282	268.2	316.9	396.8	547.5
		27	27.4	28.7	30.0	328	34.3	35.8	37.3	38.9	40.5	42.1	45.0	474	49.3	512	532	552	57.4	59.6	61.9	643	66.8	69.4	72.1	100	780	544	87.9	91.7	95.6	666		114.6	120.4	126.8	133.8	141.7	120.6	200	1873	2051	227.9	258.9	304.4	378.4	5172
		8	26.4	21.7	22	318	33.2	34.7	362	37.7	30.3	608	979	1	48.0	667	51.8	633	86.0	582	60.4	62.8	66.3	67.8	705	13.3	222	100	18	80.7	83.6	87.8		112.2	117.8	124.0	130.8	322	141	0.01	0.00	2 8	500	250.0		3009	488.4
		52	26.3	38.6	n 90	30.7	32.1	33.5	85.0	38.6	38.1	39.7	4.4	6 75	48.7	48.6	50.5	52.5	54.6	8.8	69.0	61.3	63.8	66.3	8.9	1.1	1 20	0.12	84.2	87.8	91.6	8.7	8 8		115.2	121.3		223	100	2 2						-	
# Large	Wells	Positive	•		N 6	. 4		ø	7	60		9	5 5	<u>1</u> 42	1		┢			19	_				54	+	88					8 1			8			g :			4 9		1 4				

Total Coliform and *E. coli* Bench Sheet

Missouri State University Ozarks Environmental and Water Resources Institute

Analyst:					Pro	ject:		_ Date analyzed:													
Incubator Data:	:										0	Data reviewed by:									
Start Day/Time:				Star	t Temp	erature (°C):															
End Day/Time:				End	Tempe	erature (ºC):															
					()	l Coliform yellow)		<i>E. coli</i> (yellow & <u>fluorescent</u>)													
Sample Data		Pos	e Well itive unt	Pos	l Well itive unt	Most Probable Number Coliform	Conf	5% idence imit	Pos	e Well itive unt	Pos	l Well itive unt	Most Probable Number <i>E. coli</i>	Conf	5% idence mit						
Sample	Date	Repl	licate	Repl	icate	(MPN/100mL)*	_		Replicate		Repl	icate	(MPN/100mL)*	-							
Identification	Collected	Α	В	А	В	[Mean of A + B]	-	+	А	В	А	В	[Mean of A + B]	-	+						